

# INTERNATIONAL STANDARD

ISO  
7899-1

Second edition  
1998-11-15

## Water quality — Detection and enumeration of intestinal enterococci in surface and waste water —

### Part 1:

Miniaturized method (Most Probable Number)  
by inoculation in liquid medium

*Qualité de l'eau — Recherche et dénombrement des entérocoques  
intestinaux dans les eaux de surface et résiduaires —*

*Partie 1: Méthode miniaturisée (nombre le plus probable) par  
ensemencement en milieu liquide*



Reference number  
ISO 7899-1:1998(E)

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International Organization for Standardization  
Case postale 56 • CH-1211 Genève 20 • Switzerland  
Internet iso@iso.ch  
Printed in Switzerland

## Foreword

ISO (the International Organization for Standardization) is a worldwide federation of national standards bodies (ISO member bodies). The work of preparing International Standards is normally carried out through ISO technical committees. Each member body interested in a subject for which a technical committee has been established has the right to be represented on that committee. International organizations, governmental and non-governmental, in liaison with ISO, also take part in the work. ISO collaborates closely with the International Electrotechnical Commission (IEC) on all matters of electrotechnical standardization.

Draft International Standards adopted by the technical committees are circulated to the member bodies for voting. Publication as an International Standard requires approval by at least 75 % of the member bodies casting a vote.

International Standard ISO 7899-1 was prepared by Technical Committee ISO/TC 147, *Water quality*, Subcommittee SC 4, *Biological methods*.

This second edition cancels and replaces the first edition (ISO 7899-1:1984), which has been technically revised.

ISO 7899 consists of the following parts, under the general title *Water quality — Detection and enumeration of intestinal enterococci in surface and waste water*.

- *Part 1: Miniaturized method (Most Probable Number) by inoculation in liquid medium*
- *Part 2: Method by membrane filtration*

Annexes E and F form an integral part of this part of ISO 7899. Annexes A, B, C, D and G are for information only.

## Introduction

The aim of this part of ISO 7899 is to enumerate the major intestinal enterococci, namely *E. faecalis*, *E. faecium*, *E. durans* and *E. hirae*, which occur frequently in faeces of humans and homeothermic animals. Other faecal *Enterococcus* species, namely *E. avium*, *E. cecorum*, *E. columbae* and *E. gallinarum*, and *Streptococcus bovis/equinus* strains may occasionally be included, but they occur rarely in the environmental samples. Their recovery tends to be low. *Enterococcus casseliflavus* and *E. mundtii* are non-faecal species which, when present in water samples (e.g. because of influence of plant material and some industrial effluents), are enumerated as faecal enterococci. These species and other rare non-faecal species tend to produce yellow pigment on a non-selective medium. The possible interference of non-faecal *Enterococcus* species should therefore be considered in the interpretation of results.

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# Water quality — Detection and enumeration of intestinal enterococci in surface and waste water —

## Part 1:

Miniaturized method (Most Probable Number) by inoculation in liquid medium

### 1 Scope

This part of ISO 7899 specifies a miniaturized method for the detection and enumeration of major intestinal enterococci in surface and waste water by inoculation in a liquid medium. The method is applicable to all types of surface and waste waters, particularly those rich in suspended matter.

This method is not suitable for drinking water and any other type of water for which the guideline count is less than 15 per 100 ml.

### 2 Normative references

The following standards contain provisions which, through reference in this text, constitute provisions of this part of ISO 7899. At the time of publication, the editions indicated were valid. All standards are subject to revision, and parties to agreements based on this part of ISO 7899 are encouraged to investigate the possibility of applying the most recent editions of the standards indicated below. Members of IEC and ISO maintain registers of currently valid International Standards.

ISO 3951:1989, *Sampling procedures and charts for inspection by variables for percent nonconforming*.

ISO 5667-1:1980, *Water quality — Sampling — Part 1: Guidance on the design of sampling programmes*.

ISO 5667-2:1991, *Water quality — Sampling — Part 2: Guidance on sampling techniques*.

ISO 5667-3:1994, *Water quality — Sampling — Part 3: Guidance on the preservation and handling of samples*.

ISO 8199:1988, *Water quality — General guide to the enumeration of microorganisms by culture*.

ISO/IEC Guide 2:1996, *Standardization and related activities — Vocabulary*.

### 3 Definitions

For the purposes of this part of ISO 7899, the definitions given in ISO/IEC Guide 2 and the following definition apply.

#### 3.1

##### intestinal enterococci

microorganisms capable of aerobic growth at 44 °C and of hydrolysing the 4-methylumbelliferyl-β-D-glucoside (MUG), in the presence of thallium acetate, nalidixic acid and 2,3,5-triphenyltetrazolium chloride (TTC), in the liquid medium specified

## 4 Principle

The diluted sample is inoculated in a row of microtitre plate wells containing dehydrated culture medium.

The microtitre plates are examined under ultraviolet light at 366 nm in the dark after an incubation period of between 36 h and 72 h at 44 °C ± 0,5 °C. The presence of enterococci is indicated by fluorescence resulting from the hydrolysis of MUD. The results are given as Most Probable Number (MPN) per 100 ml.

## 5 Apparatus

With the exception of equipment supplied sterile, the glassware shall be sterilized in accordance with the instructions given in ISO 8199.

Usual microbiological laboratory equipment, and in particular:

**5.1 Apparatus for sterilization** by dry heat (oven) or by steam (autoclave).

**5.2 Thermostatic incubator**, regulated at 44 °C ± 0,5 °C.

**5.3 Tunnel drier or vertical laminar air flow cabinet** (preferably class II).

**5.4 UV observation chamber** (Wood's Lamp 366 nm).

**WARNING — UV light can cause irritation of skin and eyes. Use protective gloves and glasses.**

**5.5 Portable refractometer** (optional).

**5.6 pH meter**, with an accuracy of ± 0,1.

**5.7 Test tubes**, 16 mm x 160 mm and 20 mm x 200 mm, **or flasks** with similar capacity.

**5.8 Adjustable or pre-set 8-channel multipipette**, or any system suitable for measuring and distributing 200 µl per well.

**5.9 Sterile tips** for multipipette.

**5.10 Equipment for membrane filtration**, in accordance with ISO 8199, including membrane filters with a nominal pore size of 0,2 µm, for sterilization of liquid media.

**5.11 Sterile microtitre plates**, 96-well, 350 µl, flat-bottomed, nonfluorescent.

**5.12 Sterile adhesive cover strips** for sealing microtitre plates.

**5.13 Sterile Petri dishes**, 90 mm in diameter.

## 6 Sampling

Take the samples and deliver them to the laboratory in accordance with ISO 8199 and ISO 5667-1, ISO 5667-2 and ISO 5667-3.

## 7 Culture media and diluents

### 7.1 General instructions

To ensure reproducible results, prepare culture medium and diluents, using either constituents of uniform quality and chemicals of recognized analytical or a dehydrated diluent or complete medium prepared following the manufacturer's instructions. Prepare them with distilled or demineralized water, free from substances capable of inhibiting or promoting growth under the test conditions. If the media are not used immediately, preserve them in the dark at  $(5 \pm 3)^\circ\text{C}$ , for up to one month in conditions avoiding any alterations to their composition.

NOTE The use of chemicals of other grades is permissible providing they are shown to be of equivalent performance in the test.

### 7.2 Diluent

#### 7.2.1 Special Diluent (SD)

Synthetic sea salt <sup>1)</sup>	22,5 g
Bromophenol blue solution (optional)	10 ml
Demineralized or distilled water (7.2.2)	1000 ml

Sterilize in the autoclave (5.1) at  $121^\circ\text{C} \pm 3^\circ\text{C}$  for 15 min to 20 min.

The bromophenol blue solution is prepared by adding 0,04 g in 100 ml of 50 % ethanol. It is used only to colour the SD blue and avoid confusing it with demineralized or distilled water.

#### 7.2.2 Demineralized or distilled water

Water used for dilution shall be demineralized or distilled water free from substances inhibiting growth under the test conditions.

Sterilize in the autoclave (5.1) before use at  $121^\circ\text{C} \pm 3^\circ\text{C}$  for 15 min to 20 min.

### 7.3 Culture medium: MUD/SF medium

#### 7.3.1 Composition

##### 7.3.1.1 Solution A

Tryptose	40 g
$\text{KH}_2\text{PO}_4$	10 g
D(+)-galactose	2 g
Polyoxyethylenesorbitan monooleate (Tween® 80 <sup>2)</sup>	1,5 ml
Demineralized or distilled water (7.2.2)	900 ml

Add tryptose,  $\text{KH}_2\text{PO}_4$ , galactose and Tween® 80 to 900 ml of water, whilst maintaining gentle heat and magnetic stirring, then bring to the boil until completely dissolved. Allow to cool.

1) A typical analysis of a commercially available and suitable synthetic sea salt is given in annex C. Pure NaCl solutions are not suitable, as they lead to marked inhibition.

2) Tween® 80 is an example of a suitable product available commercially. This information is given for the convenience of users of this part of ISO 7899 and does not constitute an endorsement by ISO of this product.

### 7.3.1.2 Solution B

NaHCO <sub>3</sub>	4 g
Nalidixic acid	250 mg
Demineralized or distilled water (7.2.2)	50 ml

Add both chemicals to 50 ml of water, whilst maintaining gentle heat and magnetic stirring. Allow to cool.

### 7.3.1.3 Solution C

Thallium(I) acetate	2 g
2,3,5-triphenyltetrazolium chloride	0,1 g
Demineralized or distilled water (7.2.2)	50 ml

Add both chemicals to 50 ml of water, whilst maintaining gentle heat and magnetic stirring. Allow to cool.

### 7.3.1.4 Solution D

MUD (4-methylumbelliferyl- $\beta$ -D-glucoside)	150 mg
<i>N,N</i> -dimethylformamide	2 ml

**WARNING — Thallium acetate and *N,N*-dimethylformamide are toxic. Use in a chemicals fume hood.**

### 7.3.2 Preparation

Mix together solutions A+B+C+D.

Adjust the pH to 7,5 ± 0,2.

Sterilize by filtration through a membrane of average pore size 0,2 µm (5.10).

Distribute in 96-well microtitre plates (5.11) with a volume of 100 µl of media in each well (minimum capacity 350 µl) and dehydrate immediately in a tunnel drier or laminar air-flow cabinet (5.3).

The manufacturing of the medium shall meet the quality criteria given in annex E.

## 8 Procedure

### 8.1 Choice of dilutions

The number of dilutions to inoculate varies according to the presumed level of contamination of the water to be tested. Table 1 gives some examples.

**Table 1**

Origin of sample	No. of dilutions	No. of wells /dilution	Measurement limits bacteria / 100 ml
Bathing water	2	64 wells to 1/2 32 wells to 1/20	15 to 3,5x10 <sup>4</sup>
Other surface water	4	24 wells to 1/2 24 wells to 1/20 24 wells to 1/200 24 wells to 1/2 000	40 to 3,2x10 <sup>6</sup>
Waste water and treatment plants	6	16 wells to 1/2 Up to 16 wells to 1/200 000	60 to 6,7x10 <sup>8</sup>

## 8.2 Preparation of dilutions

NOTE These procedures should be performed in a biological safety cabinet, as aerosols may be created by diluting and pipetting.

### 8.2.1 Fresh and brackish (waste) water [salinity < 30 g/kg, measured with refractometer (5.5) or equivalent method]

Prepare the relevant number of sterile tubes (5.7) in a rack, according to the number of selected dilutions; add 9 ml of the special diluent (7.2.1) to each tube.

Vigorously stir the sample (see clause 6) in order to obtain a homogeneous distribution of the microorganisms and, using a sterile pipette, immediately transfer 9 ml of this homogenized sample to the first tube containing 9 ml of diluent (7.2.1) (1/2 dilution).

Using a fresh pipette, transfer 1 ml of this dilution (homogenized) to the second tube (1/20 dilution).

From the second tube (dilution 1/20 carefully homogenized) proceed, if necessary, to another 1/10 dilution giving the dilution 1/200.

Continue as above until all the dilutions have been prepared.

### 8.2.2 Sea water (salinity $\geq 30$ g/kg)

Prepare the relevant number of sterile tubes (5.7) in a rack, according to the number of selected dilutions, add 9 ml of demineralized or distilled water (7.2.2) to the first tube and 9 ml of the special diluent (7.2.1) to the other tubes.

Vigorously stir the sample (see clause 6) in order to obtain a homogeneous distribution of the microorganisms and, using a sterile pipette, immediately transfer 9 ml of this homogenized sample to the first tube containing 9 ml water (7.2.2) (1/2 dilution).

Using a fresh sterile pipette, transfer 1 ml of this dilution (homogenized) to the second tube (1/20 dilution).

From the second tube (dilution 1/20 carefully homogenized) proceed, if necessary, to another 1/10 dilution giving the following dilution (1/200).

Continue as above until all the dilutions have been prepared.

## 8.3 Inoculation and incubation of microtitre plates

### 8.3.1 Inoculation

Transfer the contents of the first tube of dilution to an empty, sterile Petri dish, of minimum diameter 90 mm.

Using a multichannel pipette (5.8) with 8 sterile tips (5.9), distribute 200  $\mu$ l into each well of a microtitre plate (5.11) corresponding to this first dilution.

For subsequent dilutions (1/20, 1/200, etc.) operate in an identical manner, changing the Petri dish and the row of 8 sterile tips between each dilution.

Alternatively, any other suitable system (5.8) may be used to distribute 200  $\mu$ l of each dilution per well in accordance with table 1.

**CAUTION — Beware of contamination via overflow from one well to another.**

### 8.3.2 Incubation

Once the microtitre plate is inoculated, cover with the disposable sterile adhesive tape (5.12) provided for this purpose.

Incubate the microtitre plate (5.2) at  $44^{\circ}\text{C} \pm 0,5^{\circ}\text{C}$  for a minimum of 36 h and a maximum of 72 h.

NOTE The microtitre plates should be handled with care, without tilting.

## 8.4 Reading of results

Place each microtitre plate, including adhesive, in the UV observation chamber (5.4).

Consider all wells in which a blue fluorescence is observed as being positive.

NOTE The reading may be carried out any time after 36 h, as the fluorescence does not alter with time.

## 9 Expression of results

### 9.1 Determination of characteristic number

For each chosen dilution, note the number of positive (+) wells.

EXAMPLE 1 : Bathing water

1/2	32 + out of 64
1/20	5 + out of 32

Record 32/5 as characteristic number

EXAMPLE 2 : Other surface water

1/2	24 + out of 24
1/20	18 + out of 24
1/200	5 + out of 24
1/2 000	1 + out of 24

Record 18/5/1 as characteristic number

EXAMPLE 3 : Waste water

1/2	16 + out of 16
1/20	16 + out of 16
1/200	12 + out of 16
1/2 000	5 + out of 16
1/20 000	0 + out of 16
1/200 000	0 + out of 16

Record 12/5/0 as characteristic number

Where three or more dilutions have been inoculated, a characteristic number of three digits, ending in 0 where possible, shall be recorded, in accordance with ISO 8199.

## 9.2 Calculation of the MPN and its confidence interval

The MPN is a statistical estimation of the density of microorganisms, assumed to correspond to a Poisson distribution in the volumes inoculated. Confidence intervals are attached to this MPN.

Software shown in annex A or B enable the calculation of the MPN of intestinal enterococci per millilitre of water for each configuration of inoculations and the confidence interval at 95 %.

EXAMPLE 1: Assuming CN is the Characteristic Number, LO the Lower Limit and UP the Upper Limit:

If CN = 32/5, the software in annex A gives 7,56 enterococci per millilitre,

[LO = 5,42 – UP = 10,54],

i.e. 756/100 ml (542 to 1054)

EXAMPLE 2 :

If CN = 18/5/1, the software in annex A gives 159,08/ml,

[LO = 101,99 – UP = 248,11]

EXAMPLE 3 :

If CN = 12/5/0, the software in annex A gives 1 724,61/ml

[LO = 1 003,98 – UP = 2 962,50]

If none of the wells is positive, express the result in the following form:

< n/100 ml

where n is the MPN for 1 positive well under the dilution conditions employed.

## 10 Test report

The test report shall include all details necessary for the complete identification of the sample, referring to the method used and the results.

The test report shall also mention any special phenomena observed during the test and any non-specified or optional operations used in the method which may have altered the results.

## 11 Performance data

Information concerning the repeatability and reproducibility of the procedure, obtained from interlaboratory tests, is given in annex D.

## Annex A

### (informative)

#### Example of software for statistical analysis of MPNs

```
10  REM ****
20  REM GENERAL PURPOSE PROGRAM FOR MPN, ITS S.E., C.I.
30  REM AND HOMOGENEITY TEST STATISTICS
40  REM ****
50  REM
60  DIM A(10,6),X2(3,9)
70  REM SET PROGRAM LIMITS
80  D9=10
90  U9=50
100 L9=0
110 A1=.0005
120 E1=85
130 REM SET CHI-SQUARED SIGNIFICANCE LEVELS
140 GOSUB 1000
150 REM READ IN RESULTS OF A DILUTION SERIES
160 GOSUB 2000
170 REM CALC AND PRINT THE MPN
180 GOSUB 3000
190 REM CALC AND PRINT S.E. OF LOG10(MPN)
200 GOSUB 4000
210 REM CALC AND PRINT 95 PERCENT C.I. FOR MPN
220 GOSUB 5000
230 REM CALC AND PRINT DEVIANCE
240 GOSUB 6000
250 STOP
1000 REM SET CHI-SQUARED SIGNIFICANCE LEVELS
1010 FOR I=1 TO 3
1020 FOR J=1 TO 9
1030 READ X2(I,J)
1040 NEXT J
1050 NEXT I
1060 REM 5 PERCENT LEVELS DF=1...9
1070 DATA 3.84, 5.99, 7.81, 9.49, 11.07
1080 DATA 12.59, 14.07, 15.51, 16.92
1090 REM 1 PERCENT LEVELS
1100 DATA 6.63, 9.21, 11.34, 13.28, 15.09
1110 DATA 16.81, 18.48, 20.09, 21.67
1120 REM .1 PERCENT LEVELS
1130 DATA 10.83, 13.81, 16.27, 18.47, 20.52
1140 DATA 22.46, 24.32, 26.12, 27.88
1150 RETURN
2000 REM READ IN RESULTS OF A DILUTION SERIES
2010 PRINT "MPN GENERAL PURPOSE PROGRAM"
2020 PRINT ****
2030 PRINT "
2040 PRINT "M.A. HURLEY AND M.E. ROSCOE"
2050 PRINT "
2060 PRINT "NUMBER OF DILUTION LEVELS.....K=";
2070 INPUT N
2080 IF N<1 THEN GOTO 2060
2090 IF N<=D9 THEN GOTO 2120
2100 PRINT "ERROR *** LEVELS EXCEED MAXIMUM"
2110 STOP
```

```
2120  S1=0
2130  FOR I=1 TO N
2140  PRINT " "
2150  PRINT "LEVEL NUMBER .....I=",I
2160  PRINT "DILUTION FACTOR.....D=";
2170  INPUT A(I,2)
2180  PRINT "SUBSAMPLE VOLUME.....V=";
2190  INPUT A(I,1)
2200  PRINT "NUMBER OF SUBSAMPLES.....N=";
2210  INPUT A(I,3)
2220  PRINT "NUMBER OF POSITIVE SUBSAMPLES..P=";
2230  INPUT A(I,4)
2240  PRINT "IS THE DATA CORRECT FOR LEVEL ";I;"(Y OR N) ";
2250  INPUT R$
2260  IF R$="Y" THEN 2280
2270  GOTO 2140
2280  A(I,5)=A(I,1)*A(I,2)
2290  A(I,6)=A(I,5)*A(I,4)
2300  S1=S1+A(I,5)*A(I,3)
2310  NEXT I
2320  RETURN
3000  REM CALCULATES AND PRINTS MPN
3010  B1=0
3020  S3=0
3030  FOR J=1 TO N
3040  E2=A(J,5)*U9
3050  IF E2<E1 GOTO 3080
3060  E2=0
3070  GOTO 3090
3080  E2=EXP(-E2)
3090  S3=S3+A(J,6)/(1-E2)
3100  NEXT J
3110  IF S3-S1>=0 THEN 3130
3120  GOTO 3200
3130  FOR I=1 TO N
3140  A(I,5)=A(I,5)*2
3150  A(I,6)=A(I,6)*2
3160  NEXT I
3170  S1=S1*2
3180  B1=B1+1
3190  GOTO 3020
3200  X3=L9
3210  X4=U9
3220  X=(X3+X4)/2
3230  S=0
3240  FOR I=1 TO N
3250  E2=A(I,5)*X
3260  IF E2<E1 GOTO 3290
3270  E2=0
3280  GOTO 3300
3290  E2=EXP(-E2)
3300  S=S+A(I,6)/(1-E2)
3310  NEXT I
3320  IF ABS(S-S1)<A1 THEN 3380
3330  IF S-S1>0 THEN 3360
3340  X4=X
3350  GOTO 3220
3360  X3=X
3370  GOTO 3220
3380  X5=X*(2^B1)
3390  PRINT " "
```

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```
3400 PRINT "MPN=";X5
3410 PRINT "FOR A SAMPLE WITH DILUTION FACTOR 1"
3420 PRINT "                                AND VOLUME 1"
3430 RETURN
3400 REM CALCS AND PRINTS S.E. OF LOG10 (MPN)
4010 S2=0
4020 FOR I=1 TO N
4030 X3=A(I,5)
4040 E2=X3*X
4050 IF E2<E1 GOTO 4080
4060 X4=0
4070 GOTO 4090
4080 X4=EXP(-E2)
4090 S3=X3*X3*A(I,3)*X4
4100 S3=S3/(1-X4)
4110 S2=S2+S3
4120 NEXT I
4130 V=1/(X*X*S2)
4140 S1=SQR(V)/LOG(10)
4150 PRINT " "
4160 PRINT "S.E. OF LOG10 (MPN) =";S1
4170 RETURN
5000 REM CALCS 95 PERCENT C.I. FOR MPN
5010 X3=LOG(X)+B1*LOG(2)
5020 S2=SQR(V)
5030 U=EXP(X3+1.96*S2)
5040 L=EXP(X3-1.96*S2)
5050 PRINT " "
5060 PRINT "95 PERCENT C.I. =";L;"TO";U
5070 RETURN
6000 REM CALCS. AND PRINTS DEVIANCE
6010 S3=0
6020 FOR I=1 TO N
6030 S4=0
6040 IF A(I,4)<=0 GOTO 6110
6050 E2=A(I,5)*X
6060 IF E2<E1 GOTO 6090
6070 E2=0
6080 GOTO 6100
6090 E2=EXP(-E2)
6100 S4=A(I,4)*LOG(A(I,4)/(A(I,3)*(1-E2)))
6110 S3=S3+S4
6120 S4=0
6130 IF A(I,4)>=A(I,3) GOTO 6160
6140 S4=A(I,3)-A(I,4)
6150 S4=S4*(LOG(S4/A(I,3))+A(I,5)*X)
6160 S3=S3+S4
6170 NEXT I
6180 D=2*S3
6190 REM CHI-SQUARED TEST OF DEVIANCE
6200 V=N-1
6210 PRINT " "
6220 PRINT "DEFIANCE =";D;"ON";V;" D.F."
6230 PRINT " "
6240 PRINT "CHI-SQUARED SIGNIFICANCE LEVELS FOR";V;" D.F."
6250 PRINT "      5 PERCENT    ";X2(1,V)
6260 PRINT "      1 PERCENT    ";X2(2,V)
6270 PRINT "      .1 PERCENT   ";X2(3,V)
6280 RETURN
7000 END
```

## Annex B (informative)

### Example of software for computation of MPNs

```
10  DIM T (20)
20  DIM M (20)
30  DIM P (20)
40  L  = 0
45  CLS : L = L + 1
50  PRINT "CALCULATION OF MPN N° ";L
60  PRINT " ....."
70  A  = 1
80  PRINT
90  INPUT "NB OF DILUTIONS";DI : PRINT : PRINT
110 PRINT
120 P  = 0 : S = : U = 0
130 FOR I = 1 TO DI
140 PRINT "DILUTION" ; I
150 INPUT "NB OF POSITIVE WELLS .." ; P(I)
155 INPUT "NB OF WELLS .....";T(I)
160 INPUT "WATER VOLUME/WELL (ML)...";M(I) : PRINT
170 P  = P(I) + P
180 S  = ((T(I) - P(I))*M(I)) + S
190 U  = (M(I) * T(I)) + U
200 NEXT I
280 K  = 1
290 NP = (P/U) * 2 ^ (K + K / 2 + K / 4 + K / 8 + K / 16 + K / 32 + K / 64 + K / 128 + K / 256 + K / 512)
300 PM = 0
310 FOR I = 1 TO DI
320 PM = PM + ((P(I) * M (I) * EXP (- M(I) * NP)) / (1 - EXP (- M(I) * NP )))
330 NEXT I
340 IF PM < S THEN 370
350 K  = K + A
360 GOTO 290
370 DT = S - PM
380 IF ABS (DT) <=.000005 GOTO 702
390 K  = K - A
400 A  = A / 10
410 GOTO 290
702 FOR I = 1 TO DI:ES = ES + P(I):NEXT I
710 FOR I = 1 TO DI
720 K (I) = (T(I) * (M(I)^2))
730 NEXT I
740 FOR I = 1 TO DI
745 IF (NP * M(I) > 88 THEN E(I) = 1.65E38:GOTO 760
750 E(I) = (EXP (M(I) * NP) - 1)
760 NEXT I
770 LL = 0
780 FOR I = 1 TO DI
790 LL = LL + (K (I) / E (I))
800 NEXT I
810 CL = (LOG (NP) - (1.96 * (1 / (NP * SQR (LL))))) )
817 CU = (LOG (NP) + (1.96 * (1 / (NP * SQR (LL))))) )
```

```
840  NP = INT (NP * 100 + .5) / 100
850  PRINT : PRINT " NPP = ";NP;" / ML"
860  PRINT : PRINT
870  PRINT "LIMITS INF=";INT ( EXP (CL)* 100 + .5) / 100;" / ML SUP=" ;INT (EXP (CU) * 100 + .5) / 100;" / ML"
880  PRINT : PRINT: INPUT "DO YOUWANT ANOTHER MPN (Y/N)?";RE$
890  IF RE$ = "N" THEN END
900  GOTO 45
```

### Comment

After display of the run number (L, line 50),

Input the number of dilutions (line 90).

For each dilution:

- display the dilution rank (line 140),
- input the number of positive wells or tubes (line 150),
- input the number of wells inoculated with this dilution (line 155),
- input the volume of water inoculated per well, in millilitres (line 160).

Calculation of the MPN according to De Man [2] (lines 280-410).

Calculation of the lower and upper limits of confidence interval according to De Man [2] (lines 770-817).

Display

- of MPN (per ml) (line 850),
- the lower limit (per ml) (line 870),
- the upper limit (per ml) (line 870),

Question: "Another run ?"

If no: END.

## Annex C

### (informative)

#### Synthetic sea salt

##### C.1 Major ion composition of a convenient ocean synthetic sea salt

Major ion	% Total weight	Ionic concentration at 34 g/kg salinity (mg/l)
Chloride (Cl <sup>-</sup> )	47,470	18 740
Sodium (Na <sup>+</sup> )	26,280	10 454
Sulfate (SO <sub>4</sub> <sup>-2</sup> )	6,602	2 631
Magnesium (Mg <sup>+2</sup> )	3,230	1 256
Calcium (Ca <sup>+2</sup> )	1,013	400
Potassium (K <sup>+</sup> )	1,015	401
Bicarbonate (HCO <sub>3</sub> <sup>-</sup> )	0,491	194
Borate (B)	0,015	6,0
Strontium (Sr <sup>+2</sup> )	0,001	7,5
SOLIDS TOTAL		34 089,50
Water (H <sub>2</sub> O)	86,11	
	13,88	
TOTAL		99,99

##### C.2 Example for preparation from defined substances

Three basic solutions are to be made as follows:

###### Solution A

CaCl <sub>2</sub> ·2H <sub>2</sub> O	83,6 g
KCl	43,5 g
SrCl <sub>2</sub> ·6H <sub>2</sub> O	0,07 g
Distilled water	to 1 000 ml

###### Solution B

NaHCO <sub>3</sub>	15,15 g
Na <sub>2</sub> B <sub>4</sub> O <sub>7</sub>	3,0 g
Distilled water	to 1 000 ml

###### Solution C

MgSO <sub>4</sub> ·7H <sub>2</sub> O	190,0 g
MgCl <sub>2</sub> ·6H <sub>2</sub> O	147,0 g
Distilled water	to 1 000 ml

The diluent is made by adding to 960 ml distilled water 10 ml of solution A, 10 ml of solution B, 20 ml of solution C, and then 14,9 g sodium chloride, mixing until completely dissolved and setting the pH to 7,5 ± 0,2. The diluent is distributed to containers of desired volumes, and sterilized by autoclaving at (121 ± 3) °C for 15 min.

## Annex D

### (informative)

#### Performance characteristics of the method

The performances of repeatability ( $r$ ) and reproducibility ( $R$ ) calculated according to ISO 5725-2 as part of interlaboratory tests have shown:

On bathing waters

(spiked samples, with 100 French laboratories, on three occasions in 1995 and 1996) (one sea water and two fresh waters, without significant difference):

i) At the level 100 intestinal enterococci / 100 ml	$r$	$\approx 3,6$
	$R$	$\approx 5,2$
ii) At the level 400 intestinal enterococci / 100 ml	$r$	$\approx 2,1$
	$R$	$\approx 3,7$

and, for information only, during tests (in 1993 and 1994) limited to nine laboratories and four samples (naturally contaminated):

b) On river waters containing

between $2,2 \times 10^3$ and $1,5 \times 10^4$ intestinal enterococci / 100 ml:	$r$	$\approx 1,5$
	$R$	$\approx 2,7$

c) On sewage waters containing

between $1,9 \times 10^4$ and $5,1 \times 10^6$ intestinal enterococci / 100 ml:	$r$	$\approx 2,6$
	$R$	$\approx 3,9$

## Annex E (normative)

### Quality criteria for manufacturing of the medium in microtitre plates

#### E.1 General

For each of the criteria which follow, a quality control shall be made on each batch of manufactured microtitre plates. The microtitre plates to be tested are taken at random or in a systematic way to constitute a sample in accordance with ISO 3951, respecting the general control level No. II of the normal control.

The threshold of positivity of a microtitre plate is defined as being the fluorescence level leading to a positive reading without ambiguity, to the eye, under a Wood light (366 nm). It shall be measured in application of the protocol of annex F.

The quality criteria to be respected are given in E.2 to E.5. The batch is rejected if any of the criteria is not respected.

#### E.2 Background noise

Absence of positive well in each microtitre plate of the sample, after inoculation with sterile special diluent and incubation of 48 h at  $(44 \pm 0,5)$  °C. The medium background noise of the sample shall be inferior to 25 % of the positivity threshold defined above and the variation coefficient shall be inferior to 10 %.

#### E.3 Average level of fluorescence

This is the geometric mean of the fluorescence signal obtained from the 96 wells of a microtitre plate inoculated uniformly with 200 µl per well of a suspension of *E. faecalis* CCM 2541 containing 500 microorganisms per ml of Special Diluent (7.2.1), and incubated for 48 h at  $(44 \pm 0,5)$  °C. The average signal so obtained shall be at least twice the threshold of positivity and the variation coefficient shall be inferior to 10 %.

#### E.4 Fertility

Fertility is calculated as the ratio of the number of germs observed with the batch of microtitre plates under test to the number of microorganisms expected with a stable reference material (target value). The level of concentration should be brought up to around the maximum of the precision of the method, that is to say about one microorganism per well (500/100 ml). The stability and the homogeneity (target value and confidence intervals) of the reference material should have been determined with one (or several) batch(es) of microtitre plates already accepted. The threshold of acceptance of the microtitre plates tested is 0,66 to 1,5 of the target value. The variation coefficient should be inferior to 10 %.

The strains to be tested are :

■ <i>E. faecium</i> WR63	Obtainable from RIVM, Bilthoven, The Netherlands
■ <i>E. faecalis</i> CCM 2541	Obtainable from the Czech Collection of Microorganisms, Brno, Czech Republic.
■ <i>E. hirae</i> CCM 2423	

The incubation period is 48 h at  $(44 \pm 0,5)$  °C.